## Review

# Significance of angiogenesis in cancer therapy

D. A. McNAMARA, J. H. HARMEY, T. N. WALSH, H. P. REDMOND\* and D. J. BOUCHIER-HAYES

Royal College of Surgeons in Ireland Department of Surgery, Beaumont Hospital, Dublin, Ireland Correspondence to: Ms D. A. McNamara, University College Cork, Department of Surgery, Cork University Hospital, Cork, Ireland

Background For most solid tumours, surgery remains the most effective primary treatment. Despite apparently curative resection, significant numbers of patients develop secondary disease due to growth of undetected micrometastases. The ability of a tumour to metastasize is related to the degree of angiogenesis it induces. In addition, micrometastases rely on new vessel formation to provide the nutrients necessary for growth. A better understanding of how tumours acquire their blood supply may lead to more effective adjuvant therapies and improve survival following surgery.

Methods A systematic review of the literature on angiogenesis between 1971 and 1997 was performed using the Medline database to ascertain current thinking on angiogenesis and its relevance in oncological surgery.

Results Angiogenesis is a physiological process subject to autocrine and paracrine regulation which has the potential to become abnormal and play a part in a number of pathological states, including cancer. Increased angiogenic stimuli in the perioperative period, associated with concomitant reduction in tumour-derived antiangiogenic factors following resection of a primary tumour, result in a permissive environment which allows micrometastases to grow.

Conclusion Recognition of the role of angiogenesis in metastatic tumour growth represents a significant development in our understanding of tumour biology. The development of antiangiogenic agents offers new promise in the treatment of malignancy. Such agents may prevent or control the development and growth of primary and metastatic tumours.

Angiogenesis is a process through which new blood vessels develop from pre-existing vessels such as capillaries and postcapillary venules<sup>1,2</sup>. It is tightly regulated by a large number of proangiogenic and antiangiogenic factors. In physiological circumstances, angiogenesis is fully controlled by an equilibrium between these factors such that little blood vessel growth occurs in the healthy adult. Accelerated angiogenesis is a normal physiological response to wound healing, inflammation, menstruation and embryonic development, but a pathological feature of conditions such as diabetic retinopathy, rheumatoid arthritis and solid tumours.

The pivotal role of angiogenesis in primary tumour growth and metastasis has been recognized for many years, although the mechanisms which control it are incompletely understood<sup>3</sup>. Growth of a tumour beyond 2-3 mm<sup>3</sup> requires development of a microvessel network to facilitate delivery of nutrients and oxygen, and removal of catabolites. Density of microvasculature has been used as an indicator of biological aggressiveness and metastatic potential in many primary tumours<sup>4</sup>, as neovascularization facilitates metastasis<sup>1,5</sup> by providing access to the circulation<sup>6,7</sup>. The degree of angiogenesis in primary breast8, prostate4 and colorectal9 carcinoma correlates with lymph node metastasis. The predictive value of microvessel density in the primary tumour remains controversial. Microvessel density has been shown to be an independent prognostic indicator in non-small cell lung cancer10. certain instances of node-negative breast cancer<sup>11,12</sup> and several other carcinomas<sup>13</sup>. However, conflicting reports exist, particularly in relation to its prognostic value in breast cancer<sup>14,15</sup>. These conflicting findings may be attributable, in part, to the heterogeneity of microvessel density within individual tumours, as well as to interlaboratory and interobserver variability in microvessel counting<sup>12</sup>.

understanding Our mechanisms of angiogenesis in metastatic disease is increasing<sup>16</sup> Acquisition of a blood supply by micrometastases at the site of implantation is crucial to tumour growth. Neovascularization is promoted by a number of proangiogenic factors of tumour and stromal origin. The clinical impression that resection of a primary tumour heralds a phase of increased metastatic growth is of particular interest, but until recently no explanation of this phenomenon existed. Recent studies have shown that the intact primary tumour can regulate growth of associated metastases, either directly or indirectly, through the production of certain antiangiogenic factors, notably angiostatin<sup>17</sup>. Surgical excision of a primary tumour removes the source of the inhibitory angiostatin and other factors, allowing angiogenesis and subsequent growth of previously dormant micrometastases<sup>18</sup>. Tumour recurrence many years after apparently successful treatment of a primary lesion is partly the result of increased angiogenesis<sup>19,20</sup>. Long-term suppression of angiogenesis may become a therapeutic option for induction of long-term remission by maintaining micrometastases in a state of dormancy, a dynamic equilibrium during which there is no net tumour growth<sup>20</sup>. The proliferating capillary endothelial cell offers a unique target for antiangiogenic therapy<sup>21</sup> as antiangiogenic strategies may reduce both the recurrence rate and the metastatic potential of solid tumours<sup>22</sup>.

# Stages of neovascularization

Newly formed capillaries are composed of two cell types, endothelial cells and pericytes. These two cells have the capacity to produce entire capillary networks. Following the transduction of signals which promote differentiation in vivo, angiogenesis progresses in four stages: activation of endothelial cells, proliferation, migration and lumen formation<sup>23</sup>.

Cytokine release is provoked by factors endogenous and exogenous to tumour cells, including local environmental factors such as hypoxia. Quiescent endothelial cells are activated by the release of cytokines from host and tumour cells (stage 1). Committed cells proliferate (stage 2), then migrate along a fibrin skeleton towards the source of the angiogenic stimulus to form cords of aligned cells (stage 3). Finally, the vascular sprout forms a lumen and the cells exit the cell cycle to a resting phase. Development of a patent lumen (stage 4) occurs through coalescence of intracellular vacuoles and is facilitated by cell-to-cell adhesive contact1.

Degradation of the extracellular matrix is an essential component of new vessel invasion. This is facilitated by alteration of the proteolytic balance. Proteolytic degradation of the extracellular matrix and fibrinolysis are two functions of pericytes<sup>24</sup>, although pericytes are also thought to contribute by production of growth factors and growth inhibitors. Cell adhesion receptors promote vascular cell migration through interaction with adhesion proteins of the extracellular matrix, such as collagen and fibronectin. The extracellular matrix also serves as a reservoir for growth factors, particularly acidic fibroblast growth factor and basic fibroblast growth factor (bFGF)<sup>25-27</sup>.

The capillary sprouts in tumours are 'leaky' as proliferating capillaries have incomplete basement membranes. In addition, vascular endothelial growth factor (VEGF; previously known as vascular permeability factor) increases permeability through the development of a series of interconnected cytoplasmic vesicles and vacuoles (known as vesical-vacuolar organelles) that maintain contact with both the luminal and abluminal surface<sup>28,29</sup>. In normal tissues this may play a regulatory role in controlling baseline microvascular permeability<sup>30</sup>, and in tumour microvasculature this feature has been linked to malignant exudates and ascites<sup>31,32</sup>.

## Regulation of angiogenesis

The degree of angiogenesis in a tumour is the result of complex interactions between tumour cells, capillary endothelial cells, pericytes and recruited immune cells with products of these cells acting in both an autocrine and paracrine fashion<sup>33</sup>. It is normally subject to tight physiological control through a balance of proangiogenic and antiangiogenic factors. These factors allow a phase of rapid proliferation to facilitate wound healing but maintain quiescence in the mature microvasculature<sup>5</sup>. Increased production of positive angiogenic factors is 'necessary but not sufficient'34 for induction of the angiogenic phenotype; negative regulators must also be decreased<sup>18,35,36</sup>. Imbalance between angiogenic promoters and inhibitors produces the intense angiogenesis which is characteristic of many pathological processes, including diabetic retinopathy<sup>37–39</sup>, rheumatoid arthritis<sup>40</sup>, endometriosis<sup>41</sup> and malignant tumours<sup>29,42</sup>.

Factors influencing angiogenesis are derived both from tumour cells and infiltrating cells, such as macrophages and fibroblasts7. Through their secretory products, activated macrophages can influence each phase of the angiogenic process<sup>43–45</sup>. The process of macrophage activation is mediated in part by hypoxia<sup>46</sup>. The large number of macrophages present in a variety of angiogenesis-associated pathological states is indicative of their influence. It is known, for example, that numerous macrophages are present in the first phase of tumour growth. Macrophage density is directly proportional to rate of tumour growth in breast carcinoma<sup>47</sup>. Furthermore, tumour-associated macrophages cause marked augmentation of tumour neovascularization<sup>40</sup> and correlate directly with prognosis in this disease<sup>48</sup>. Other immune cells are also significant, as neutrophilia is associated with a poorer prognosis in breast cancer<sup>49</sup>.

# Proangiogenic factors (Table 1)

Vascular endothelial growth factor

VEGF is the most potent directly acting angiogenic protein known<sup>50</sup>. It is a diffusible endothelial cell-specific

Table 1 Proangiogenic substance:

Substance	Reference	
Vascular endothelial growth	Marme <sup>30</sup>	1996
factor	man the second	1000
Basic fibroblast growth	Rifkin and Moscatelli <sup>74</sup>	1989
factor (FGF-2) Acidic fibroblast growth	Jouanneau et al. 177	1995
factor (FGF-1)	Jouanneau er ar.	1997
Platelet-derived endothelial	Takahashi et al.7	1996
cell growth factor	rusumism crun.	+2311
Platelet activating factor	Camussi et al.178	1995
Fransforming growth factor β1	Pepper et al. 102	1993
Transforming growth factor a	Gleave et al.179	1993
Insulin-like growth factor	Nicosia et al. 180	1994
Epidermal growth factor	Goldman et al.181	1993
Interleukin 1	Fan et al.182	1993
Interleukin 4	Wojta et al. 183	1993
Interleukin 6	Sunderkotter et al.43	1994
Interleukin 8	Smith et al.184	1994
Interleukin 15	Angiolillo et al. 185	1997
Angiogenin	Moenner et al.186	1994
Vascular integrins $\alpha v_1 \beta_2$ and $\alpha v_1 \beta_5$	Varner and Cheresh <sup>1</sup>	1996
Fibronectin	Bitterman et al.26	1983
Fibrin	Tsopanoglau et al.107	1993
Tumour necrosis factor  a (in vivo)	Frater-Schroder et al.111	1987
Endotoxin (lipopolysaccharide)	Mattsby-Baltzer et al. 112	1994
Matrix metalloproteinases	Cornelius et al.188	1995
Scatter factor (hepatocyte growth factor)	Rosen et al. 189	1993
Hyaluron fragments	Cockerill et al.23	1995
Granulocyte-macrophage colony stimulating factor	Bikfalvi and Han <sup>190</sup>	1994
Platelet activating factor	Bussolino et al,191	1995
Proliferin	Jackson et al.192	1994
Substance P	Fan et al.182	1993
Lactate	Magee et al.193	1994
Prostaglandin E <sub>1</sub> , E <sub>2</sub>	Ben-Av et al.194	1995
Urokinase plasminogen activator	Hildenbrand et al.,195	1995
Tenascin	Canfield and Schor <sup>106</sup>	1995
Nitric oxide	Ziche et al.197	1994
Erucamide	Mitchell et al. 108	1996

mitogen and angiogenic factor, which also increases vascular permeability<sup>51,52</sup>. It binds to two endothelial cell tyrosine kinase receptors, the fms-like tyrosine kinase (flt) and the kinase domain receptor (KDR)<sup>53,54</sup>.

Tumour-derived VEGF plays an important role in the paracrine stimulation of angiogenesis but it also appears to have an autocrine stimulatory effect on tumour cells<sup>55</sup>, particularly in response to hypoxia<sup>56</sup>. Hypoxia stimulates angiogenesis in a number of sites, including endothelial cells, retinal pericytes<sup>57</sup>, myocardium<sup>58</sup> and solid tumours<sup>59</sup>. VEGF activity is potentiated by oxygen deprivation<sup>60-62</sup>, mediated in part by adenosine<sup>57,63</sup> through upregulation of the VEGF endothelial cell receptor KDR<sup>64</sup>. Wild-type p53, a tumour suppressor gene, inhibits proliferation of both normal and transformed cells<sup>65,66</sup>, and also regulates VEGF production. Mutations in this gene abolish such control. There appear to be two regulatory VEGF pathways: an oncogenic one (v-src) that enhances VEGF production and a tumour suppressor (p53) signal that suppresses VEGF<sup>50</sup>. Mutations of the ras oncogene cause VEGF upregulation in a colon cancer model with resulting increased angiogenesis<sup>67</sup>. H-ras oncogene mutation also activates angiogenesis through upregulation VEGF and matrix metalloproteinase (MMP) bioactivity, while downregulating activity of tissue inhibitors of MMP<sup>68</sup>.

Tumour cell lines which express minimal constitutive and hypoxia-inducible VEGF (e.g. human melanoma SK-MEL-2 cells) produce small, poorly vascularized tumours. Overexpression of VEGF results in dramatically increased angiogenesis and subsequent tumour expansion<sup>69</sup>. Interruption of the VEGF receptor paracrine pathway in glioma tumours inhibits angiogenesis, leading to reduced numbers of new vessels, a higher degree of necrosis and a reduction in size of glioma tumours<sup>70</sup>. Expression of VEGF and its receptor KDR is higher in metastatic than in non-metastatic colonic neoplasms, and correlates directly with the extent of neovascularization and degree of proliferation<sup>9</sup>. There is a direct correlation between serum VEGF concentration and breast cancer stage<sup>71</sup>.

Soluble recombinant flt receptor binds to free VEGF and inhibits its mitogenic affinity for endothelial cells<sup>72</sup>. Anti-VEGF antibodies have been used as a new therapeutic modality for the prevention of retinal ischaemia-induced neovascularization of the iris, confirming that the *in vitro* effects of VEGF inhibition are reproducible *in vivo*<sup>73</sup>.

## Basic fibroblast growth factor

This is a 16-kDa protein which is one of a family of fibroblast growth factors<sup>74</sup>. Capillary endothelial cells themselves produce and release bFGF which acts in an autocrine fashion independently, and synergistically with VEGF<sup>75</sup>, as an endothelial cell mitogen<sup>76</sup>. It binds intensely to the extracellular matrix, but is normally confined to its cell of origin as it lacks the signal peptide thought to be necessary for secretion<sup>50</sup>. During the malignant evolution of fibrosarcoma, neovascularization is associated with a switch to the ability to export bFGF from cells<sup>77</sup>. It also stimulates endothelial and vascular smooth muscle cell motility<sup>78,79</sup> especially after arterial trauma<sup>80</sup>. Heparin released by recruited mast cells amplifies its effect<sup>5</sup>. Its systemic administration results in an increased rate of tumour growth and increased vascularity<sup>81</sup>. Binding by bFGF is inhibited by suramin, an antiparasitic agent<sup>82</sup>. It has been shown that blockade of

bFGF receptors results in suppression of solid tumour growth<sup>83</sup>.

## Acidic fibroblast growth factor

This is a heparin binding site-specific endothelial cell mitogen<sup>84,85</sup> which promotes angiogenesis and wound healing<sup>86,87</sup>. It is expressed constitutively in normal as well as in tumour cells, but appears to be inhibited by an angiogenesis suppressor gene which is downregulated during tumorigenesis<sup>34</sup>. Like bFGF, it is found in many normal cells but exported only in tumour cells.

## Platelet-derived endothelial cell growth factor

This proangiogenic cytokine is found in platelets<sup>88</sup> and released during the blood clotting cascade, as well as being produced by vascular smooth muscle cells89. It is an endothelial cell mitogen but is not a classical growth factor, because direct contact with a cell-surface receptor is not required for its mitogenic capability<sup>90</sup>. It promotes endothelial cell migration<sup>91</sup> and may facilitate endothelial differentiation<sup>92</sup>. Platelet-derived endothelial cell growth factor (PD-ECGF) expression is known to be modulated by the tumour microenvironment<sup>93</sup>. It is homologous to thymidine phosphorylase<sup>94</sup>, an enzyme which catalyses the reversible breakdown of thymidine to thymine and deoxyribose-1-phosphate. This enzymatic activity may contribute to its angiogenic activity, as products of deoxyribose-1-phosphate are known to be angiogenic, although the mechanism through which PD-ECGF exerts is angiogenic effect is not yet fully understood 95,96.

### Transforming growth factor $\beta$

Transforming growth factor (TGF)  $\beta$ , which is produced by macrophages and activated platelets, exists in three human isoforms,  $\beta 1$ ,  $\beta 2$  and  $\beta 3$ . All act as chemotactic agents for macrophages and are indirectly angiogenic *in vivo* <sup>97-101</sup>, although TGF  $\beta 1$  appears most potent in this regard<sup>100</sup>. TGF  $\beta 1$  facilitates wound healing through accelerated collagen deposition and maturation<sup>101</sup>. It induces matrix production and alters integrin production to facilitate neovascularization. It acts in a dose-dependent manner, with low concentrations exhibiting stimulatory and high concentrations inhibitory effects<sup>102</sup>. Its absence results in poor vascular integrity and reduced remodelling<sup>103</sup>. TGF  $\beta 3$  appears to have a greater role in matrix remodelling, with a resultant decrease in scar formation<sup>100</sup>.

#### Epithelial growth factor

Epidermal growth factor (EGF) is a mitogenic growth factor. EGF receptors belong to a group of protooncogenes, including c-erbB2, which are overexpressed in a number of human tumours<sup>104</sup>. EGF is secreted by tumour-associated macrophages in patients with carcinoma of the breast<sup>105</sup>.

#### Integrins

Blood vessels involved in angiogenesis express adhesion proteins, known as integrins, including von Willebrand factor, fibronectin, fibrin and vascular integrin  $\alpha v_1 \beta_3$  and  $\alpha v_1 \beta_5$ . They facilitate cell-cell and cell-matrix interactions <sup>106</sup>, thereby playing a crucial role in endothelial

cell chemotaxis. Adhesion proteins may be selectively inhibited by monoclonal antibodies with the resulting decrease in angiogenesis inducing apoptosis of endothelial cells<sup>34</sup>. Vascular integrin  $\alpha v_1 \beta_3$  is selectively expressed on growing vessels, where it suppresses p53 activity while increasing the bcl2:bax ratio with a net proangiogenic and antiapoptotic effect<sup>107</sup>. Its inhibition by monoclonal antibodies in a melanoma model results in reduced angiogenesis, suggesting that the new tumour vasculature grows in a self-perpetuating manner 108.

#### Tumour necrosis factor a

Tumour necrosis factor (TNF) a is a macrophage-derived angiogenic factor which is considered to be the principal mediator of tumour cell cytotoxicity and cytostasis 109. It has a broad spectrum of biological activities including both stimulatory and inhibitory effects on target cells which are dose related 40,110. It induces granulocytemacrophage colony stimulating factor production, has procoagulant activity and increases the adherence of human endothelial cells. Its proangiogenic activity was initially postulated to be induced through an inflammatory mechanism secondary to its cytotoxic effects<sup>111</sup>. It is now believed that TNF-α is a potent mediator of angiogenesis at low concentrations, independent of inflammation 110.

#### Endotoxin

Endotoxin (lipopolysaccharide; LPS) is a ubiquitous molecule derived from the cell wall of most Gramnegative and some Gram-positive bacteria as well as existing in a free form in the environment. It is a proangiogenic molecule because of its ability to elicit production of a wide spectrum of host-effector molecules, such as VEGF, bFGF, TGF- $\beta$ , TNF, interleukin (IL) 1 and IL-6, by macrophages and other cell populations 112-114. Recent work from the authors' laboratory shows that LPS introduced at the time of open surgery in a tumourbearing murine model increases serum VEGF concentration<sup>115</sup>.

## Antiangiogenic factors (Table 2)

#### **Thrombospondin**

Thrombospondin is a glycoprotein secreted by many cells, including endothelial and epithelial cells, fibroblasts, smooth muscle cells, monocytes and macrophages<sup>116</sup>. It modulates endothelial cell adhesiveness, motility and proteolytic activity by sequestering angiogenesis inducers<sup>117</sup>. While it is a potent inhibitor of angiogenesis, it is downregulated during tumorigenesis<sup>5,118,119</sup>. In fibroblasts, wild-type p53 inhibits angiogenesis through upregulation of thrombospondin<sup>33</sup>, an effect which is abolished by p53 mutations. Paradoxically, thrombospondin enhances the angiogenic effect of LPS and bFGF stimuli in inflammatory states 120. It is speculated that thrombospondin 1 downregulates VEGF expression through an as yet unknown mechanism<sup>48</sup>.

## Angiostatin

Angiostatin is a 38-kDa internal fragment of plasminogen<sup>121</sup> which inhibits endothelial cell proliferation, angiogenesis and tumour growth. It blocks the catalytic site of the enzyme that cleaves plasminogen, thereby

Table 2 Antiangiogenic substances

Substance	Reference	Year
Thrombospondin	Canfield and Schor <sup>196</sup>	1995
Angiostatin	O'Reilly et al.121	1994
Endostatin	O'Reilly et al. 123	1997
Suramin	Takano et al.170	1994
Interferon $\alpha$ and $\beta$	Stout et al.168	1993
Interferon y	Kobayashi et al.199	1995
Interferon $\mu$ inducible protein 10	Angiolillo et al.200	1996
Interleukin 12	Voest et al. 169	1995
Tamoxifen	Donovan et al.32	1997
Thalidomide	D'Amato et al.158	1994
Linomide	Vukanovic and Isaacs <sup>201</sup>	1995
Fumagillin analogues (AGM-1470)	Klauber et al. 160	1997
Captopril	Volpert et al.202	1996
Platelet factor 4	Strieter et al. <sup>203</sup>	1995
Antisense VEGF RNA, VEGF blocking antibody and VEGF receptor kinase inhibitors	Millauer et al. <sup>204</sup>	1996
Transforming growth factor $\beta$ 1	Pepper et al.102	1993
Proliferin-related protein	Jackson et al. 192	1994
2-methoxyoestradiol	Fotsis et al. 205	1994
Tissue inhibitor of metalloproteinase	Tagikawa et al. <sup>206</sup>	1990
Retinoids	Pienta et al.207	1993
Tumour necrosis factor $\alpha$ (in vitro)	Frater-Schroder et al. <sup>111</sup>	1987
Group B Streptococcus toxin CM 101	Thurman et al. <sup>208</sup>	1996
Pentosan polysulphate	Nguyen et al.209	1993
Lavendustin A	Hu and Fan <sup>210</sup>	1995
Dexamethasone	Hori et al. <sup>211</sup>	1996
Medroxyprogesterone acetate	Yamamoto et al.212	1994
Korean misletoe extract (Viscum album coloratum)	Yoon et al. <sup>213</sup>	1995
Indomethacin	Tarnawski et al.214	1989
Diclofenac	Lala et al.215	1997

VEGF, vascular endothelial growth factor

preventing the matrix remodelling required angiogenesis<sup>117</sup>. It selectively prevents endothelial cells responding to angiogenic stimuli resulting in dormancy of micrometastases<sup>122</sup>. Systemic administration has been shown to induce regression of primary tumours of breast, colon and prostate in a murine model, providing the first example of dormancy therapy in which tumours undergo regression through blockade of angiogenesis<sup>17</sup>.

#### Endostatin

Endostatin is a protein fragment of collagen XVIII and was first isolated from a murine haemangioendothelioma cell line<sup>123</sup>. It acts in a similar way to angiostatin, blocking collagenase and impeding matrix remodelling. specifically inhibits endothelial cell proliferation. Recombinant endostatin inhibits angiogenesis, growth of metastases and growth of primary tumours<sup>123</sup>. Endostatintreated tumours have a proliferation rate which is the same as that of untreated tumours, but an apoptotic rate which is sevenfold greater, providing a further example of dormancy therapy.

## Tumour angiogenesis

Malignant transformation is a cumulative process requiring loss of control of the cell cycle and a shift in the

balance of proangiogenic and antiangiogenic factors. Induction of angiogenesis is a local event which is specific to activated hyperplastic cells and which procedes overt tumour formation<sup>124</sup>. For example, angiogenesis is a marker of premalignant transformation in benign breast disease 125,126. Vascular endothelial cells in malignant breast tumours express tissue factor, a potent procoagulant, whereas vascular endothelial cells in benign tumours do not<sup>127</sup>. Cell proliferation and malignant transformation represent a switch to an angiogenic phenotype, but only in the latter is control of the cell cycle lost. Furthermore. only those hyperplastic cells which acquire angiogenic capacity undergo malignant transformation 124,126. One model involves a tumour suppressor gene encoding the angiogenesis inhibitory factor thrombospondin, which is downregulated when cells undergo malignant transformation, allowing the angiogenic phase to commence<sup>118</sup>. This downregulation is under the control of the p53 tumour suppressor gene<sup>33</sup>. Loss of suppressor gene sequences<sup>128</sup> leads to failure of inhibitory pathways with resultant tumour growth<sup>110</sup>.

Numerous parallels between wound healing and tumour growth exist, suggesting that tumours act as 'obligate parasites' which have developed the capacity to pre-empt and subvert the wound healing response of the host as a means to acquire the stroma they need to grow and expand<sup>129</sup>. Normal cells involved in wound healing, such as endothelial cells and fibroblasts, develop features similar to those of malignant cells<sup>130</sup>. These features include increased proteolysis<sup>131</sup>, increased motility<sup>132</sup> and proliferation. It is postulated that the invasive, proliferative behaviour of transformed cells may result from a functional failure of signals that usually regulate the activated behaviour of normal cells in a wound<sup>129</sup>.

#### Angiogenesis and tumour metastasis

Tumour cells must detach from a primary tumour to initiate the 'metastatic cascade'<sup>19,133</sup>. Angiogenesis is a necessary precursor of metastasis as new proliferating capillaries have incomplete basement membranes and are 'leaky', facilitating penetrance by tumour cells<sup>134,135</sup>. These tumour cells must evade immune surveillance in the circulation and also at the site of secondary development, where they cross endothelium, degrade basement membrane, implant, proliferate and establish their own capillary network<sup>19</sup>. Angiogenesis is necessary at both the beginning and the end of the metastatic cascade. There is resultant metastatic inefficiency as only a small number of tumour cells which leave the primary tumour successfully implant at distant sites<sup>136,137</sup>.

## Effect of surgery on angiogenesis

Clinical experience and experimental evidence support the concept that the healing wound is an immunologically privileged site for metastases<sup>25,106,138,139</sup>. The cytokine environment that ensues in the first 24 h following surgical trauma influences the healing process for the subsequent 3 weeks<sup>101</sup>. Tumour growth is enhanced in healing wounds, but not in surrounding normal tissues. It has been shown that when tumour cells reach a healing colonic anastomosis or laparotomy wound within 2 h of its formation, the probability of a tumour cell leading to a metastatic deposit is increased 1000-fold compared with normal tissue<sup>140</sup>. Evidence showing an increased hazard rate for relapse or death in the 3 years following surgery

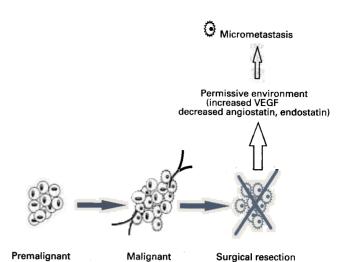


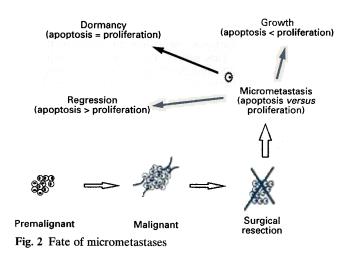
Fig. 1 Effect of surgery on micrometastases. VEGF, vascular endothelial growth factor; LPS, lipopolysaccharide

for breast cancer, and a second peak a number of years later, supports the theory that micrometastases with a variable rate of growth are present at the time of diagnosis<sup>141</sup>. Surgery and subsequent healing may contribute to the genesis of recurrent disease through promotion of a proangiogenic climate which alters the dynamic equilibrium that would otherwise maintain micrometastases in a state of dormancy<sup>20,142</sup> (Fig. 1).

VEGF may be one of the primary links between these observations, as it is necessary to allow wound healing but may also foster growth of micrometastases through its potent proangiogenic activity. Raised circulating levels of VEGF have been identified in the early postoperative period (J. H. Harmey, unpublished observation). This may facilitate the growth and metastasis of circulating or intravasated dormant tumour cells.

Endotoxin induces angiogenesis in a dose-dependent fashion at biologically relevant doses<sup>112</sup>. Significant translocation of endotoxin into the peritoneal cavity and systemic circulation occurs following open surgery or airlaparoscopy<sup>143</sup>. insufflation It directly increases macrophage production of VEGF, a potent angiogenic factor regulating both vessel growth and permeability, an effect which also appears to extend to tumour cells 115,144. Surgical trauma is known to increase leucocyte numbers and activity<sup>145</sup> and to promote tumour establishment and growth146. The authors' preliminary observations indicate that endotoxin introduced at the time of surgery may encourage the growth of metastases by suppression of host antitumour immune function and stimulation of proangiogenic factor release. The result is a permissive environment for tumour growth and angiogenesis.

It has been shown that the primary tumour produces circulating inhibitors of angiogenesis, called angiostatin and endostatin, which act specifically at the level of endothelial cell proliferation<sup>121</sup>. Within 5 days of removal of the primary tumour, angiostatin and endostatin disappear from the circulation and angiogenesis within dormant micrometastases becomes possible. This heralds a phase of rapid growth of previously dormant micrometastases<sup>18</sup>. Dormancy is normally maintained in tumour cells by means of a high replication rate and a correspondingly high apoptotic rate (Fig. 2). There is no change in the replication rate of neovascularized



metastases; however, the apoptotic rate is significantly decreased with resultant net metastatic growth<sup>20</sup>

## Therapeutic opportunities (Fig. 3)

The aim of antiangiogenic treatment is to reduce and maintain tumours as small relatively dormant clusters of cells which have low metastatic potential, are more susceptible to cell-mediated immunological attack3, and which may be more vulnerable to chemotherapy147 and radiotherapy<sup>148,149</sup>. Tumour growth is associated with increased interstitial pressure due to leaky tumour vessels<sup>150</sup> in the absence of adequate lymphatic vessels<sup>151</sup>. This causes vascular compression and eventual central necrosis. As a result, tumours do not outgrow their blood supply, they compress it152. Antiangiogenic therapy paradoxically increases delivery of chemotherapy to a tumour<sup>147</sup> by reducing interstitial pressure<sup>152</sup>. Inhibition of angiogenesis results predominantly in a cytostatic, not a cytotoxic, effect<sup>22,153–155</sup>, although a recent study suggests that antiangiogenic therapy may lead to tumour regression156.

Successful antiangiogenic strategies have incorporated a wide range of antiangiogenic agents<sup>121,157–163</sup>. Prolonged courses of interferon  $\alpha$  induce regression of human haemangiomatous disease 164. Systemic administration of angiostatin induces regression of primary murine tumours

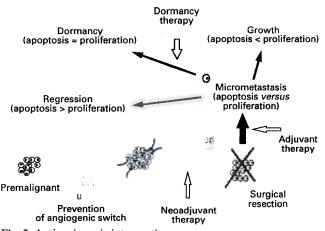


Fig. 3 Antiangiogenic interventions

of breast, colon and prostate<sup>17</sup>. Recombinant endostatin inhibits angiogenesis, the growth of metastases and the growth of primary tumours<sup>123</sup>. In vivo inhibition of angiogenesis using an animal model of pancreatic carcinoma has been shown to reduce tumour growth 165.

# Mechanisms of action

Antiangiogenic therapies may act either directly, by specifically inhibiting endothelial cell proliferation, or indirectly, by altering the cytokine microenvironment which controls angiogenesis. Tumours consist of a 'twocompartment system' in which tumour cells and endothelial cells co-exist, and produce growth factors which have paracrine effects on each other<sup>33</sup>. Inhibition of endothelial cell growth reduces production of tumourpromoting substances and inhibits tumour expansion. Inadequate tumour vascularization may result in tumour cell apoptosis, a genetically programmed form of cell death<sup>36</sup>. Dormant micrometastases are biologically active with a rate of apoptosis equal to their rate of cell proliferation 122,166. As a result, no net growth of the metastasis occurs. Antiangiogenic therapies promote and maintain perpetual dormancy by increasing the apoptotic rate within the metastasis, hence the appellation 'dormancy therapy'.

## Direct antiangiogenic agents

Angiostatin and endostatin are the only direct inhibitors of angiogenesis identified to date, and both have been shown to reduce tumour growth 121,123. Systemic administration of endostatin, which directly inhibits endothelial cell mitogenesis, has been suggested as an ideal 'dormancy therapy' 123. More recently, it has been reported that repeated cycles of endostatin can prevent tumour recurrence, possibly through the induction of an antiangiogenic phenotype, without leading to problems of drug resistance which characterize many chemotherapeutic agents<sup>156</sup>. Hanahan<sup>167</sup> has suggested that direct inhibitors of angiogenesis may be subdivided into three groups. First, pure angiogenesis inhibitors, which inhibit new vessel growth but have no effect on existing tumour vessels. Second, tumour vessel toxins, which damage existing tumour vasculature and, finally, dual action agents, which combine these two effects. It appears that endostatin is a member of the last group.

#### Indirect antiangiogenic agents

Indirectly acting antiangiogenic therapies influence the microenvironment which regulates tumour angiogenesis and so indirectly influence endothelial cell behaviour. The majority of antiangiogenic agents currently identified belong to this group. Each tumour has a unique complex microenvironment dictated both by tumour histology and site of implantation. Additionally, within any tumour, heterogeneity arises because of variations in stromal density, tumour differentiation, hypoxia and local cytokines, among other factors. As a result, each tumour is exposed to different proangiogenic and antiangiogenic stimuli which ultimately influence net angiogenesis. It is therefore unlikely that therapeutic manipulation of a single cytokine will be sufficient to regulate tumour angiogenesis adequately. Intertumour variability suggests that therapeutic regimens based on cytokine manipulation must be custom-designed for each particular tumour type.

Maximum clinical gain may potentially be achieved by a combination of antiangiogenic interventions which influence different aspects of angiogenesis. For example, the fumagillin analogue AGM-1470 reduces endothelial cell proliferation<sup>157</sup>. Interferon  $\alpha$  interferes with endothelial cell migration<sup>168</sup> and IL-12 inhibits angiogenesis mediated by interferon  $\gamma^{169}$ . In contrast, suramin inhibits bFGF receptor binding in addition to a number of direct effects on endothelial cells<sup>170</sup>, while tamoxifen inhibits VEGF-mediated endothelial cell proliferation and migration<sup>171</sup>. Direct manipulation of endothelial cell proliferation combined with alteration of the cytokine microenvironment may prove more effective than either approach alone.

Multimodality therapy, in which antiangiogenic strategies are combined with chemotherapeutic agents or radiotherapy, is known to be more effective than monotherapy, probably because of the combined effects on both the tumour and vascular compartments<sup>147–149,172</sup>.

Optimal methods and timing of administration of antiangiogenic therapies have yet to be determined. Unfocused interruption of angiogenesis may potentially interfere with wound healing, menstruation embryogenesis. Inhibitors of endothelial-specific growth factors (e.g. VEGF) have a theoretical advantage over inhibition of growth factors with more widespread targets (e.g. bFGF) because undesired non-specific interruption of mitogenesis is avoided. However, it is likely that novel methods of site-specific drug delivery, developed to administer experimental proangiogenic therapies in the treatment of ischaemic heart disease, will also be applied to the delivery of antiangiogenic agents and may help to reduce unwanted systemic effects<sup>56</sup>. Substantial expression of VEGF receptors is predominantly limited to the tumour vasculature. Interference with this system using monoclonal antibodies, dominant negative receptor or antisense oligonucleotides against VEGF messenger RNA offer the potential of tumour-specific inhibition of angiogenesis and subsequent metastatic growth 173. A similar approach could be pursued for other cytokines. Developments in the field of gene therapy<sup>174</sup> may be harnessed to produce long-term low-grade antiangiogenic substances which maintain micrometastases in a state of dormancy.

The dosing schedule for biological antiangiogenic agents is likely to differ significantly from that for pharmaceutical agents<sup>18</sup>. Clinical experience interferon  $\alpha$  in the treatment of life-threatening haemangiomas has shown that very prolonged courses of daily therapy at optimal doses are essential for vessel involution<sup>175</sup>. Timing of antiangiogenic therapy in relation other therapeutic interventions also requires clarification. For example, excision of a primary tumour results in decreased levels of the inhibitory factors angiostatin and endostatin<sup>18</sup>, and raised circulating levels of VEGF. In the postoperative setting, this is likely to facilitate the growth and metastasis of circulating or dormant tumour cells. Therefore, manipulation of tumour angiogenesis in the perioperative period provides a tempting therapeutic opportunity. The effect of impaired angiogenesis on wound healing, however, must be clarified. A balance between the neovascularization necessary for wound healing and inhibition of neoplastic microvessel formation must be achieved.

Variability in mechanisms and control of angiogenesis at different stages in the natural history of cancer may exist. A premalignant phase is recognized for many

tumours. This is also a preangiogenic phase and development of an antiangiogenic strategy which prevents the onset of angiogenesis could prevent progression to a malignant state 176. This would require both identification of premalignant cells and development of an appropriate prophylactic antiangiogenic regimen. This is likely to differ substantially from the therapeutic approach required for primary tumours or micrometastases. For example, VEGF messenger RNA expression develops late in the course of tumour progression, is not a feature of premalignancy<sup>28</sup> and its manipulation is, therefore, unlikely to be of benefit in this situation. Timing of prophylactic therapy will also be critical, as angiogenesis is probably not involved in the interim stage of micrometastasis following shedding from the primary tumour, when micrometastases are distributed in organs as either single cells or small clusters with ready access to a blood supply<sup>20</sup>. None the less, inhibition of angiogenesis is the most promising new therapy in the treatment of malignancy. Complete understanding of the factors regulating angiogenesis and the interactions between these factors should lead to valuable new interventions. These may be combined with surgery and other modalities to improve the treatment of many solid tumours.

### References

- 1 Varner J, Cheresh DA. Tumour angiogenesis and the role of vascular cell integrim  $\alpha_v \beta_3$ . In: DeVita VT, Hellman S, Rosenberg SA, eds. *Important Advances in Oncology*. Philadelphia, Pennsylvania: Lippincott, 1996: 69–87.
- 2 Noden DM. Embryonic origins and assembly of blood vessels. Am Rev Respir Dis 1989; 140: 1097–103.
- 3 Folkman J. Tumor angiogenesis: therapeutic implications. *N Engl J Med* 1971; 285: 1182–6.
- 4 Weidner N, Carroll PR, Flax J, Blumenfeld W, Folkman J. Tumor angiogenesis correlates with metastasis in invasive prostate carcinoma. Am J Pathol 1993; 143: 401-9.
- 5 Folkman J, Shing Y. Angiogenesis. J Biol Chem 1992; 267: 10 931-4.
- 6 Liotta LA, Kleinerman J, Saidel GM. Quantitative relationships of intravascular tumor cells, tumor vessels, and pulmonary metastases following tumor implantation. *Cancer Res* 1974; 34: 997–1004.
- 7 Takahashi Y, Bucana CD, Liu W, Yoneda J, Kitadai Y, Cleary KR et al. Platelet-derived endothelial cell growth factor in human colon cancer angiogenesis: role of infiltrating cells. J Nat Cancer Inst 1996; 88: 1146-51.
- 8 Weidner N, Semple JP, Welch WR, Folkman J. Tumor angiogenesis and metastasis-correlation in invasive breast carcinoma. *N Engl J Med* 1991; 324: 1–8.
- 9 Takahashi Y, Kitadai Y, Bucana CD, Cleary KR, Ellis LM. Expression of vascular endothelial growth factor and its receptor, KDR, correlates with vascularity, metastasis, and proliferation of human colon cancer. *Cancer Res* 1995; 55: 3964-8.
- 10 Macchiarini P, Fontanini G, Hardin MJ, Squartini F, Angelletti CA. Relation of neovascularisation to metastasis of non-small-cell lung cancer. *Lancet* 1992; 340: 145-6.
- 11 Toi M, Inada K, Suzuki H, Tominaga T. Tumor angiogenesis in breast cancer: its importance as a prognostic indicator and the association with vascular endothelial growth factor expression. *Breast Cancer Res Treat* 1995; 36: 193-204.
- 12 Heimann R, Ferguson D, Powers C, Recant WM, Weichselbaum RR, Hellman S. Angiogenesis as a predictor of long-term survival for patients with node-negative breast cancer. *J Natl Cancer Inst* 1996: 88: 1764-9.
- 13 Weidner N. Intratumor microvessel density as a prognostic factor in cancer. *Am J Pathol* 1995; 147: 9–19.

- 14 Costello P, McCann A, Carney DN, Dervan PA. Prognostic significance of microvessel density in lymph node negative breast carcinoma. Hum Pathol 1995; 26: 1181-4.
- 15 Van Hoef ME, Knox WF, Dhesi SS, Howell A, Schor AM. Assessment of tumor vascularity as a prognostic factor in lymph node negative invasive breast cancer. Eur J Cancer 1993; **29A**: 1141–5.
- 16 Bikfalvi A. Significance of angiogenesis in tumour progression and metastasis. Eut J Cancer 1995; 31A: 1101-4.
- O'Reilly MS, Holmgren L, Chen C, Folkman J. Angiostatin induces and sustains dormancy of human primary tumors in mice. Nat Med 1996; 2: 689-92.
- 18 Folkman J. Angiogenesis in cancer, vascular, rheumatoid and other disease. Nat Med 1995: 1: 27-31.
- 19 Jiang WG, Puntis MCA, Hallett MB. Molecular and cellular basis of cancer invasion and metastasis: implications for treatment. Br J Surg 1994; 81: 1576-90.
- 20 Uhr JW, Scheuermann RH, Street NE, Vitetta ES. Cancer dormancy: opportunities for new therapeutic approaches. Nat Med 1997; 3: 505-9.
- 21 Walker RA. Angiogenesis and breast cancer prognosis a continuing issue. J Pathol 1996; 180: 6-7.
- 22 Tanaka T, Manome Y, Wen P, Kufe DW, Fine HA. Viral vector-mediated transduction of a modified platelet factor 4 cDNA inhibits angiogenesis and tumor growth. Nat Med 1997; 3: 437-42.
- 23 Cockerill GW, Gamble JR, Vadas MA. Angiogenesis: models and modulators. Int Rev Cytol 1995; 159: 113-60.
- Nehls V, Schuchardt E, Drenckhahn D. The effect of fibroblasts, vascular smooth muscle cells, and pericytes on sprout formation of endothelial cells in a fibrin gel angiogenesis system. Microvasc Res 1994; 48: 349-63.
- Lee JY, Murphy SM, Scanlon EF. Effect of trauma on implantation of metastatic tumor in bone in mice. J Surg Oncol 1994; 56: 178-84.
- 26 Bitterman PB, Rennard SI, Adelberg S, Crystal RG. Role of fibronectin as a growth factor for fibroblasts. J Cell Biol 1983; 97: 1925-32.
- 27 Fava RA, McClure DB. Fibronectin-associated transforming growth factor. J Cell Physiol 1987; 131: 184-9.
- Senger DR, Van de Water L, Brown LF, Nagy JA, Yeo KT, Yeo TK et al. Vascular permeability factor (VPF, VEGF) in tumor biology. Cancer Metastasis Rev 1993; 12: 303-24.
- Cornali E, Zietz C, Benelli R, Weninger W, Masiello L, Breier G et al. Vascular endothelial growth factor regulates angiogenesis and vascular permeability in Kaposi's sarcoma. Am J Pathol 1996; 149: 1851-69.
  30 Berse B, Brown LF, Van de Water L, Dvorak HF, Senger
- DR. Vascular permeability factor (vascular endothelial growth factor) gene is expressed differentially in normal tissues, macrophages, and tumors. Mol Biol Cell 1992; 3:
- 31 Nagy JA, Herzberg KT, Masse EM, Zientara GP, Dvorak HF. Exchange of macromolecules between plasma and peritoneal cavity in ascites tumor-bearing, normal, and serotonin-injected mice. Cancer Res 1989; 49: 5448-58.
- 32 Donovan D, Harmey JH, Redmond HP, Bouchier-Hayes D. Ascites revisited: a novel role for tamoxifen. Eur J Surg Oncol 1997; 23: 570.
- 33 Folkman J. Tumor angiogenesis and tissue factor. Nat Med 1996; 2: 167-8.
- 34 Folkman J. Angiogenesis inhibitors generated by tumors. Mol Med 1995; 1: 120-2.
- 35 Dameron KM, Volpert OV, Tainsky MA, Bouck N. Control of angiogenesis in fibroblasts by p53 regulation of thrombospondin-1. *Science* 1994; **265**: 1582–4.
- 36 Hanahan D, Folkman J. Patterns and emerging mechanisms of the angiogenic switch during tumorigenesis. Cell 1996; 86: 353-64.
- 37 Aiello LP, Avery RL, Arrigg BA, Keyt BA, Jampel HD, Shah ST et al. Vascular endothelial growth factor in ocular fluid of patients with diabetic retinopathy and other retinal disorders. *N Engl J Med* 1994; 331: 1480-7. 38 Miller JW, Adamis AP, Shima DT, D'Amore PA, Moulton

- RS, O'Reilly MS et al. Vascular endothelial growth factor/ vascular permeability factor is temporally and spatially correlated with ocular angiogenesis in a primate model. Am J Pathol 1994; 145: 574-84.
- 39 Murata T, Ishibashi T, Khalil A, Hata Y, Yoshikawa H, Inomata H. Vascular endothelial growth factor plays a role in hyperpermeability of diabetic retinal vessels. Opthalmic Res 1995; 27: 48-52.
- 40 Polverini PJ. Macrophage-induced angiogenesis: a review. In: Sorg C, ed. Macrophage-derived Cell Regulatory Factors.
- Basel: Karger, 1989: 54-73.

  41 McLaren J, Prentice A, Charnock-Jones DS, Millican SA, Muller KH, Sharkey AM et al. Vascular endothelial growth factor is produced by peritoneal fluid macrophages in endometriosis and is regulated by ovarian steroids. J Clin Invest 1996; 98: 482-9.
- 42 Samoto K, Ikezaki K, Ono M, Shono T, Kohno K, Kuwano M et al. Expression of vascular endothelial growth factor and its possible relation with neovascularization in human brain tumors. Cancer Res 1995; 55: 1189-93.
- 43 Sunderkotter C, Steinbrink K, Goebeler M, Bhardwaj R, Sorg C. Macrophages and angiogenesis. J Leukoc Biol 1994; 55: 410-22
- 44 Harmey J, Dimitriadis E, Redmond HP, Bouchier-Hayes D. Macrophage production of vascular endothelial growth factor (VEGF) is differentially regulated by hypoxia and transforming growth factor  $\beta$ -1. Ann Surg Oncol 1998; 5: 271-8.
- 45 Harmey JH, Toomey D, Osborne DH, Redmond HP, Bouchier-Hayes D. Elevated vascular endothelial growth factor in breast cancer: a proposed mechanism. Surgical Forum 1996; XLVII: 522-4.
- 46 Knighton DR, Hunt TK, Scheuenstuhl H, Halliday BJ, Werb Z, Bands MJ. Oxygen tension regulates the expression of angiogenesis factor by macrophages. Science 1983; 221: 1283-5.
- 47 van Netten JP, Ashmed BJ, Cavers D, Fletcher C, Thornton IG, Antonsen BL et al. 'Macrophages' and their putative significance in human breast cancer. Br J Cancer 1992; 66: 220-1 (Letter).
- 48 Mantovani A. Tumor-associated macrophages in neoplastic progression: a paradigm for the in vivo function of chemokines. Lab Invest 1994; 71: 5-16.
- 49 Welch DR, Schissel DJ, Howley RP, Aeed PA. Tumorelicited polymorphonuclear cells, in contrast to 'normal' circulating polymorphonuclear cells, stimulate invasive and metastatic potentials of rat mammary adenocarcinoma cells.
- Proc Natl Acad Sci U S A 1989; 86: 5859-63.
  50 Mukhopadhyay D, Tsiokas L, Sukhatme VP. Wild-type p53 and v-Src exert opposing influences on human vascular endothelial growth factor gene expression. Cancer Res 1995;
- 51 Ferrara N. The role of vascular endothelial growth factor in pathological angiogenesis. Breast Cancer Res Treat 1995; 36: 127-37.
- 52 Marme D. Tumor angiogenesis: the pivotal role of vascular endothelial growth factor. World J Urol 1996; 14: 166-74.
- 53 de Vries C, Escobedo JA, Ueno H, Houck N, Ferrara N, Williams LT. The fms-like tyrosine kinase, a receptor for vascular endothelial growth factor. Science 1992; 255:
- 54 Terman BI, Dougher-Vermazen NM, Carrion ME, Dimitrov D, Armellino DC, Gospodarowicz D et al. Identification of the KDR tyrosine kinase as a receptor for vascular endothelial growth factor.. Biochem Biophys Res Commun 1992; 187: 1579-86.
- 55 Liu B, Earl HM, Baban D, Shoaibi M, Fabra A, Kerr DJ et al. Melanoma cell lines express VEGF receptor KDR and respond to exogenously added VEGF. Biochem Biophys Res Commun 1995; 217: 721-7.
- 56 Namiki A, Brogi E, Kearney M, Kim EA, Wu T, Couffinhal T et al. Hypoxia induces vascular endothelial growth factor in cultured human endothelial cells. J Biol Chem 1995; 270: 31 189-95.

- 57 Takagi H, King GL, Robinson GS, Ferrara N, Aiello LP. Adenosine mediates hypoxic induction of vascular endothelial growth factor in retinal pericytes and endothelial cells. *Invest Opthalmol Vis Sci* 1996; 37: 2165-76.
- 58 Ware JA, Simons M. Angiogenesis in ischemic heart disease. *Nat Med* 1997; 3: 158-64.
- 59 Shweiki D, Itin A, Soffer D, Keshet E. Vascular endothelial cell growth factor induced by hypoxia may mediate hypoxiainitiated angiogenesis. *Nature* 1992; 359: 843-5.
- 60 Minchenko A, Salaceda S, Bauer T, Caro J. Hypoxia regulatory elements of the human vascular endothelial growth factor gene. *Cell Mol Biol Res* 1994; 40: 35-9.
- 61 Levy AP, Levy NS, Wegner S, Goldberg MA. Transcriptional regulation of the rat vascular endothelial growth factor gene by hypoxia. *J Biol Chem* 1995; 270: 13 333-40.
- 62 Grone HJ, Simon M, Grone EF. Expression of vascular endothelial growth factor in renal vascular disease and renal allografts. *J Pathol* 1995; 177: 259-67.
- 63 Hashimoto E, Kage K, Ogita T, Nakaoka T, Matsuoka R, Kira Y. Adenosine as an endogenous mediator of hypoxia for induction of vascular endothelial growth factor mRNA in U-937 cells. Biochem Biophys Res Commun 1994; 204: 318-24.
- 64 Brogi E, Schatterman G, Wu T, Kim EA, Varticovski L, Keyt B et al. Hypoxia-induced paracrine regulation of vascular endothelial growth factor receptor expression. J Clin Invest 1996; 97: 469-76.
- 65 Finlay CA, Hinds PW, Levine AJ. The p53 proto-oncogene can act as a suppressor of transformation. Cell 1989; 57: 1083-93.
- 66 Levine AJ, Momand J, Finlay CA. The *p53* tumour suppressor gene. *Nature* 1991; 351: 453-6.
- 67 Rak J, Mitsuhashi Y, Bayko L, Filmus J, Shirasawa D, Sasazuki T *et al.* Mutant *ras* oncogenes upregulate VEGF/VPF expression: implications for induction and inhibition of tumor angiogenesis. *Cancer Res* 1995; 55: 4575–80.
- 68 Arbiser JL, Moses MA, Fernandez CA, Ghiso N, Cao Y, Klauber N et al. Oncogenic H-ras stimulates tumor angiogenesis by two distinct pathways. Proc Nat Acad Sci U S A 1997; 94: 861-6.
- 69 Claffey KP, Brown LF, del Aguila LF, Tognazzi K, Yeo KT, Manseau EJ et al. Expression of vascular permeability factor/vascular endothelial growth factor by melanoma cells increases tumor growth, angiogenesis, and experimental metastasis. Cancer Res 1996; 56: 172–81.
- 70 Saleh M, Stacker SA, Wilks AF. Inhibition of growth of C6 glioma cells in vivo by expression of antisense vascular endothelial growth factor sequence. Cancer Res 1996; 56: 393-401.
- 71 Donovan D, Harmey JH, Toomey D, Osborne DH, Redmond HP, Bouchier-Hayes DJ. Transforming growth factor β-1 regulation of vascular endothelial growth factor production by breast cancer cells. Ann Surg Oncol 1997; 4: 621-7.
- 72 Kendall RL, Thomas KA. Inhibition of vascular endothelial cell growth factor activity by an endogenously encoded soluble receptor. *Proc Natl Acad Sci U S A*: 1993; 90: 10705-9.
- 73 Adamis AP, Shima DT, Tolentino MJ, Gragoudas ES, Ferrara N, Folkman J et al. Inhibition of vascular endothelial growth factor prevents retinal-ischemia induced iris neovascularization in a nonhuman primate. Arch Opthalmol 1996; 114: 66-71.
- 74 Rifkin DB, Moscatelli D. Recent developments in the cell biology of basic fibroblast growth factor. J Cell Biol 1989; 109: 1-6.
- 75 Asahara T, Bauters C, Zheng LP, Takeshita S, Bunting S, Ferrara N et al. Synergistic effect of vascular endothelial growth factor and basic fibroblast growth factor on angiogenesis in vivo. Circulation 1995; 92 (Suppl II): II 365-71.
- 76 Schweigerer L, Neufeld G, Friedman J, Abraham JA, Fiddes JC, Gospodarowicz D. Capillary endothelial cells

- express basic fibroblast growth factor, a mitogen that promotes their own growth. *Nature* 1987: 325: 257-9.
- 77 Kandel J, Bossy-Wetzel E, Radvanyi F, Klagsbrun M, Folkman J, Hanahan D. Neovascularization is associated with a switch to the export development of fibroscarcoma. *Cell* 1991; 66: 1095-104.
- 78 Biro S, Yu Z-X, Fu Y-M, Smale G, Sasse J, Sanchez J et al. Expression and subcellular distribution of basic fibroblast growth factor are regulated during migration of endothelial cells. Circ Res 1994; 74: 485-94.
- 79 Jackson CL, Reidy MA. Basic fibroblast growth factor: its role in the control of smooth muscle cell migration. Am J Pathol 1993; 143: 1024-31.
- 80 Lindner V, Reidy MA. Proliferation of smooth muscle after vascular injury is inhibited by an antibody against basic fibroblast growth factor. *Proc Natl Acad Sci USA* 1991; 88: 3739-43
- 81 Gross JL, Herblin WF, Dvorak BA, Czerniak P, Diamond MD, Sun T et al. Effects of modulation of basic fibroblast growth factor on tumor growth in vivo. J Natl Cancer Inst 1993; 85: 121-31.
- 82 Danesi R, Del Bianchi S, Soldani P, Campagni A, La Rocca RV, Myers CE et al. Suramin inhibits bFGF-induced endothelial cell proliferation and angiogenesis in the chick chorioallantoic membrane. Br J Cancer 1993; 68: 932–8.
- 83 Hori A, Sasada R, Matsutani E, Naito K, Sakura Y, Fujita T. Suppression of solid tumor growth by immunoneutralizing monoclonal antibody against human basic fibroblast growth factor. Cancer Res 1991; 51: 6180-4.
- 84 Folkman J, Klagsbrun M. Angiogenic factors. Science 1987; 235: 442-7.
- 85 Thompson JA, Anderson KD, DiPietro JM, Zwiebel JA, Zametta M, Anderson WF et al. Site-directed neovessel formation in vivo. Science 1988; 241: 1349-52.
- 86 Klagsbrun M. The fibroblast growth factor family: structural and biological properties. *Progress in Growth Factor Research* 1989; 1: 207–35.
- 87 Thomas KA, Gimenez-Gallego G, Di Salvo J, Linemeyer D, Kely L, Menke J et al. Structure and activities of acidic fibroblast growth factor. In: Rifkin DB, Klagbrun M, eds. Current Communications in Molecular Biology: Angiogenesis Mechanisms and Pathobiology. Cold Spring Harbor, New York: Cold Spring Harbor Laboratories, 1987: 9-12.
- 88 Miyazono K, Okabe T, Urabe A, Takaku F, Heldin CH. Purification and properties of an endothelial cell growth factor from human platelets. *J Biol Chem* 1987; 262: 4098-103.
- 89 Usuki K, Heldin N-E, Miyazono K, Ishikawa F, Takaku F, Westermark B et al. Production of platelet-derived endothelial cell growth factor by normal and transformed human cells in culture. Proc Natl Acad Sci USA 1989; 86: 7427-31.
- 90 Finnis C, Dodsworth N, Pollitt CE, Carr G, Sleep D. Thymidine phosphorylase activity of platelet-derived endothelial cell growth factor is responsible for endothelial cell mitogenicity. *Eur J Biochem* 1993; 212: 201–10.
- 91 Risau W, Drexler H, Mironov V, Smits A, Siegbahn A, Funa K et al. Platelet-derived growth factor is angiogenic in vivo. Growth Factors 1992; 7: 261-6.
- 92 Klagsbrun M, D'Amore PA. Regulators of angiogenesis. *Annu Rev Physiol* 1991; 53: 217–39.
- 93 Griffiths L, Dachs GU, Bicknell R, Harris AL, Stratford IJ. The influence of oxygen tension and pH on the expression of platelet-derived endothelial cell growth factor/thymidine phosphorylase in human breast tumor cells grown in vitro and in vivo. Cancer Res 1997; \$7: 570-72.
- 94 Furukawa T, Yoshimura A, Sumizawa T, Haraguchi M, Akiyama S, Fukui K et al. Angiogenic factor. Nature 1992; 356: 668 (Letter)
- 356: 668 (Letter).
  95 O'Brien TS, Harris AL. Angiogenesis in urological malignancy. Br J Urol 1995; 76: 675–82.
- 96 Griffiths L, Stratford IJ. Platelet-derived endothelial cell growth factor thymidine phosphorylase in tumour growth and response to therapy. *Br J Cancer* 1997; 76: 689–93.

- 97 Thompson AM, Kerr DJ, Steel CM. Transforming growth factor  $\beta$  1 is implicated in the failure of tamoxifen therapy in human breast cancer. Br J Cancer 1991; 63: 609-14.
- 98 Roberts AB, Sporn MB, Assoian RK, Smith JM, Roche NS, Wakefield LM et al. Transforming growth factor type  $\beta$ : rapid induction of fibrosis and angiogenesis in vivo and stimulation of collagen formation in vitro. Proc Natl Acad Sci USA 1986; 83: 4167-71.
- Sporn MB, Roberts AB, Wakefield LM, de Crombrugghe B. Some recent advances in the chemistry and biology of transforming growth factor-β. J Cell Biol 1987; 105: 1039-45.
- 100 Shah M, Foreman DM, Ferguson MW. Neutralisation of TGF  $\beta$ -1 and TGF  $\beta$ -2 or exogenous addition of TGF- $\beta$  3 to cutaneous rat wounds reduces scarring. J Cell Sci 1995; 108: 985-1002.
- 101 Pierce GF, Tarpley JE, Yanagihara D, Mustoe TA, Fox GM, Thomason A. Platelet-derived growth factor (BB homodimer), transforming growth factor- $\beta 1$ , and basic fibroblast growth factor in dermal wound healing. Neovessel and matrix formation and cessation of repair. Am J Pathol 1992; 140: 1375-88
- 102 Pepper MS, Vassali JD, Orci L, Montesano R. Biphasic effect of transforming growth factor-β 1 on *in vitro* angiogenesis. *Exp Cell Res* 1993; 204: 356–63.
- 103 Folkman J, D'Amore PA. Blood vessel formation: what is its molecular basis? Cell 1996; 87: 1153-5
- 104 Baillie CT, Winslet MC, Bradley NJ. Tumour vasculature a potential therapeutic target. Br J Cancer 1995; 72: 257-67.
- 105 O'Sullivan C, Lewis CE, Harris AL, McGee JO. Secretion of epidermal growth factor by macrophages associated with breast carcinoma. Lancet 1993; 342: 148-9.
- 106 Murthy MS, Scanlon EF, Silverman RH, Goodheart CR, Goldschmidt RA, Jelachich ML. The role of fibronectin in tumor implantation at surgical sites. Clin Exp Metastasis 1993; 11: 159-73.
- 107 Stromblad S, Becker JC, Yebra M, Brooks PC, Cheresh DA. Suppression of p53 activity and p21WAF1/CIP1 expression by vascular cell integrin  $\alpha_v \beta_3$  during angiogenesis. J Clin Invest 1996; 98: 426-33.
- 108 Brooks PC, Clark RAF, Cheresh DA. Requirement of vascular integrin alpha <sub>γβ3</sub> for angiogenesis. *Science* 1994; 264: 569-71
- 109 Le J, Vilcek J. Biology of disease. Tumour necrosis factor and interleukin-1: cytokines with multiple overlapping biological activities. Lab Invest 1987; 56: 234-48.
- 110 Leibovich SJ, Polverini PJ, Shepard HM, Wiseman DM, Shively V, Nuseir N. Macrophage-induced angiogenesis is mediated by tumour necrosis factor-alpha. Nature 1987; 329:
- 111 Frater-Schroder M, Risau W, Hallmann R, Gautschi P, Bohlen P. Tumor necrosis factor type alpha, a potent inhibitor of endothelial cell growth in vitro, is angiogenic in vivo. Proc Natl Acad Sci USA 1987; 84: 5277-81.
- 112 Mattsby-Baltzer I, Jokobsson A, Sorbo J, Norrby K. Endotoxin is angiogenic. *Int J Exp Pathol* 1994; 75: 191-6.
- 113 Watson RW, Redmond HP, Bouchier-Hayes D. Role of endotoxin in mononuclear phagocyte-mediated inflammatory responses. *J Leukoc Biol* 1994; 56: 95–103.
- 114 Li WW, Grayson G, Folkman J, D'Amore PA. Sustainedrelease endotoxin. A model for inducing corneal neovascularization. Invest Ophthalmol Vis Sci 1991; 32: 2906-11.
- 115 Pidgeon G, Harmey JH, Kay E, Da Costa M, Redmond HP, Bouchier-Hayes D. The effect of surgery and endotoxin on tumour growth and angiogenesis. Ir J Med Sci 1997; 166: 8.
- 116 Raugi GJ, Mumby SM, Abott-Brown D, Bornstein P. Thrombospondin: synthesis and secretion by cells in culture. J Cell Biol 1982; 95: 351-4.
- 117 Bussolino F, Mantovani A, Persico G. Molecular mechanisms of blood vessel formation. Trends Biochem Sci 1997; 22: 251-6,
- 118 Rastinejad F, Polverini PJ, Bouck NP. Regulation of the activity of a new inhibitor of angiogenesis by a cancer

- suppressor gene. Cell 1989; 56: 345-55.
- 119 Bouck N. Tumour angiogenesis: the role of oncogenes and tumour suppressor genes. Cancer Cells 1990; 2: 179-85.
- 120 BenEzra D, Griffin BW, Maftzir G, Aharonov Thrombospondin and in vivo angiogenesis induced by basic fibroblast growth factor or lipopolysaccharide. Invest Ophthalmol Vis Sci 1993; 34: 3601-8.
- 121 O'Reilly M, Holmgren L, Shing Y, Chen C, Rosenthal RA, Moses M et al. Angiostatin: a novel angiogenesis inhibitor that mediates the suppression of metastases by a Lewis lung carcinoma. Cell 1994; 79: 315-28.
- 122 Holmgren L, O'Reilly MS, Folkman J. Dormancy of micrometastases: balanced proliferation and apoptosis in the presence of angiogenesis suppression. Nat Med 1995; 1: 149-53.
- 123 O'Reilly MS, Boehm T, Shing Y, Fukai N, Vasios G, Lane WS et al. Endostatin: an endogenous inhibitor of angiogenesis and tumor growth. Cell 1997; 88: 277-85.
- 124 Folkman J, Watson K, Ingber D, Hanahan D. Induction of angiogenesis during the transition from hyperplasia. Nature 1989; 339: 58-61.
- 125 Brem SS, Jensen HM, Gullino PM. Angiogenesis as a marker for preneoplastic lesions of the human breast. Cancer 1978; 41: 239-44.
- 126 Brem SS, Gullino PM, Medina D. Angiogenesis: a marker for neoplastic transformation of mammary papillary hyperplasia. Science 1977; 195: 880-2.
- 127 Contrino J, Hair G, Kreutzer D, Rickles FR. In situ detection of tissue factor in vascular endothelial cells: correlation with the malignant phenotype of human breast disease. Nat Med 1996; 2: 209-15.
- 128 Sager R. Tumour suppressor genes: the puzzle and the promise. Science 1989; 246: 1406-12.
- Whalen GF. Solid tumours and wounds: transformed cells misunderstood as injured tissue? Lancet 1990; 336: 1489-92.
- 130 Dvorák HF. Tumors: wounds that do not heal. Similarities between tumor stroma generation and wound healing. N Engl J Med 1986; 315: 1650+9.
- 131 Kalebic T, Garbisa S, Glaser B, Liotta LA. Basement membrane collagen: degradation by migrating endothelial
- cells. Science 1983; 221: 281-3.
  132 Postlewaithe AE, Keski-Oja J, Moses HL, Kang AH. Stimulation of the chemotactic migration of human fibroblasts by transforming growth factor beta. J Exp Med 1987; **165**: 251-6.
- 133 Liotta LA, Stracke ML. Tumor invasion and metastases: biochemical mechanisms. Cancer Treat Res 1988; 40: 223-38.
- 134 Liotta LA, Saidel G, Kleinerman J. The significance of hematogenous tumor cell clumps in the metastatic process. Cancer Res 1976; 36: 889-94.
- 135 Nagy JA, Brown LF, Senger DR, Lanir N, Van de Water L, Dvorak AM et al. Pathogenesis of tumor stroma generation: a critical role for leaky blood vessels and fibrin deposition. Biochim Biophys Acta 1989; 948: 305-26.
- 136 Fidler IJ, Gerstein DM, Hart IR. The biology of cancer invasion and metastasis. Adv Cancer Res 1978; 28: 149–250.
- 137 Nicholson G. Cancer metastasis. Sci Am 1979; 240: 66-76.
- 138 Ammirati M, Rao LN, Murthy MS, Buchmann T, Goldschmidt RA, Scanlon EF. Partial nephrectomy in mice with milliwatt carbon dioxide laser and its influence on experimental metastasis. J Surg Oncol 1989; 41: 153-9.
- 139 Murthy MS, Goldschmidt RA, Rao LN, Ammirati M, Buchmann T, Scanlon EF. Influence of surgical trauma on experimental metastasis. Cancer 1989; 64: 2035-44.
- 140 Skipper D, Jeffrey MJ, Cooper AJ, Alexander P, Taylor I. Enhanced growth of tumour cells in healing colonic anastomoses and laparotomy wounds. Int J Colorectal Dis
- 141 Baum M, Badwe RA. Does surgery influence the natural history of breast cancer? In: Wise L, Johnson J Jr, eds. Breast Cancer: Controversies in Management. Armonk, New York: Futura Publishing, 1994: 61-9.
- 142 Baum M. Breast cancer: a challenge to the contemporary paradigm. Acta Oncol 1996; 35(Suppl 8): 3-6.

- 143 Watson RW, Redmond HP, McCarthy J, Burke PE, Bouchier-Hayes D. Exposure of the peritoneal cavity to air regulates early inflammatory responses to surgery in a murine model. Br J Surg 1995; 82: 1060-5.
- 144 McNamara DA, Pidgeon G, Harmey J, Redmond HP, Walsh TN, Bouchier-Hayes ĎJ. Exposure lipopolysaccharide elicits vascular endothelial growth factor production by macrophage Gastroenterology 1998; (in press). macrophages and tumour
- 145 Redmond HP, Watson RW, Houghton T, Condron C, Watson RG, Bouchier-Hayes D. Immune function in patients undergoing open vs laparoscopic cholecystectomy. Arch Surg 1994; 129: 1240-6.
- 146 Da Costa ML, Redmond HP, Bouchier-Hayes DJ. Increased tumor establishment and growth after laparotomy vs laparoscopy. Arch Surg 1996; 131: 1003 (Letter).
- 147 Teicher BA, Holden SA, Ara G, Sotomayor EA, Huang ZD, Chen YN et al. Potentiation of cytotoxic cancer therapies by TNP-470 alone and with other anti-angiogenic agents. Int J Cancer 1994; 57: 920-5
- 148 Teicher BA, Holden SA, Ara G, Northey D. Response of the FSaII fibrosarcoma to antiangiogenic modulators plus cytotoxic agents. Anticancer Res 1993; 13(6A): 2101-6.
- 149 Teicher BA, Ara G, Menon K, Schaub RG. In vivo studies with interleukin-12 alone and in combination with monocyte colony-stimulating factor and/or fractionated radiation treatment. Int J Cancer 1996; 65: 80-4.
- 150 Dvorak HF, Nagy JA, Dvorak JT, Dvorak AM. Identification of and characterization of the blood vessels of solid tumors that are leaky to circulating macromolecules. Am J Pathol 1988; 133: 95-109.
- 151 Jain RK. Physiological barriers to delivery of monoclonal antibodies and other macromolecules in tumors. Cancer Res 1990; 50: 814-19s.
- 152 Folkman J. The influence of angiogenesis research on management of patients with breast cancer. Breast Cancer Res Treat 1995; 36: 109-18.
- 153 Folkman J. Successful treatment of an angiogenic disease. N Engl J Med 1989; 320: 1211-12.
- 154 Folkman J. Seminars in Medicine of the Beth Israel Hospital, Boston. Clinical applications of research on angiogenesis. N Engl J Med 1995; 333: 1757-63.
- 155 Harris AL. Antiangiogenesis for cancer therapy. Lancet 1997; **349**(Suppl II): 13–15. 156 Boehm T, Folkman J, Browder T, O'Reilly MS.
- Antiangiogenic therapy of experimental cancer does not induce acquired drug resistance. *Nature* 1997; 390: 404–7. Ingber D, Fujita T, Kishimoto S, Sudo K, Kanamura T,
- Brem H et al. Synthetic analogues of fumagillin that inhibit angiogenesis and suppress tumour growth. Nature 1990; 348: 555**–**7.
- 158 D'Amato RJ, Loughnan MS, Flynn E, Folkman J. Thalidomide is an inhibitor of angiogenesis. Proc Natl Acad Sci USA 1994; 91: 4082-5.
- Malone TE, Gray GS, Petro J, Hunt AJ, Donner AL, Bauer SI et al. Inhibition of angiogenesis by recombinant human platelet factor-4 and related peptides. Science 1990; 247: 77-9.
- 160 Klauber N, Rohan RM, Flynn E, D'Amato RJ. Critical components of the female reproductive pathway are suppressed by the angiogenesis inhibitor AGM-1470. Nat Med 1997; 3: 443-6.
- 161 Effert PJ, Gastl G, Strohmeyer T. Current and future strategies to block tumor angiogenesis, invasion, and metastasis. World J Urol 1996; 14: 131-40.
- Brem H, Folkman J. Analysis of experim antiangiogenic therapy. J Pediatr Surg 1993; 28: 445-51. experimental 162 Brem H,
- 163 Brem H, Gresser I, Grosfeld J, Folkman J. The combination of antiangiogenic agents to inhibit primary tumor growth and metastasis. *J Pediatr Surg* 1993; 28: 1253-7. 164 White CW, Sondheimer HM, Crouch EC, Wilson H, Fan
- LL. Treatment of puylmonary hemangiomatosis with recombinant interferon alfa-2a. N Engl J Med 1989; 320: 1197-200.

- 165 Egawa S, Tsutsumi M, Konishi Y, Kobari M, Matsuno S, Nagasaki K et al. The role of angiogenesis in the tumor growth of Syrian hamster pancreatic cancer cell line HPD-NR. Gastroenterology 1995; 108: 1526-33.
- 166 Wu J. Apoptosis and angiogenesis: two promising tumor markers in breast cancer. Anticancer Res 1996; 16: 2233-9.
- 167 Hanahan DA. A flanking attack on cancer. Nat Med 1998; 4:
- 168 Stout AJ, Gresser I, Thompson WD. Inhibition of wound healing in mice by local interferon alpha/beta injection. Int J Exp Pathol 1993; 74: 79-85.
- Voest EE, Kenyon BM, O'Reilly MS, Truitt G, D'Amato RJ, Folkman J. Inhibition of angiogenesis in vivo by interleukin 12. J Nat Cancer Inst 1995; 87: 581-6.
- 170 Takano S, Gatley S, Neville ME, Herblin WF, Gross JL, Engelhard H et al. Suramin, an anticancer and angiosuppressive agent, inhibits endothelial cell binding of basic fibroblast growth factor, migration, proliferation, and induction of urokinase-type plasminogen activator. Cancer Res 1994; 54; 2654-60.
- 171 McNamara DA, Harmey J, Wang JH, Donovan D, Kaye E, Walsh TN et al. Tamoxifen is antiangiogenic in vitro and attenuates VEGF-mediated angiogenesis in vivo. Ir J Med Sci 1998; (in press).
- 172 Sledge GW Jr. Implications of the new biology for therapy
- in breast cancer. Semin Oncol 1996; 23(1Suppl 2): 76-81.

  173 Martiny-Baron G, Marme D. VEGF-mediated tumour angiogenesis: a new target for cancer therapy. Curr Opin Biotechnol 1996; 6: 675-80.
- 174 Blau H, Khavari P. Gene therapy: progress, problems, prospects. Nat Med 1997; 3: 612-13.
- 175 Ezekowitz RAB, Mulliken JB, Folkman J. Interferon alfa 2a for life-threatening hemangiomas in infancy. N Engl J Med 1992; **326**: 1456-63.
- 176 Klein G. The approaching era of the tumor suppressor genes. Science 1987; 238: 1539-45.
  177 Jouanneau J, Moens G, Montesano R, Thiery JP. FGF-1 but not FGF-4 secreted by carcinoma cells promotes in vitro and in vivo angiogenesis and rapid tumor proliferation. Growth Factors 1995; 12: 37-47.
- 178 Camussi G, Montrucchio G, Lupia E, De Martino A, Perona L, Arese M et al. Platelet-activating factor directly stimulates in vitro migration of endothelial cells and promotes in vivo angiogenesis by a heparin-dependent mechanism. J Immunol 1995; 154: 6492-501.
- 179 Gleave ME, Hsieh JT, Wu HC, Hong SJ, Zhau HE, Guthrie PD et al. Epidermal growth factor receptor-mediated autocrine and paracrine stimulation of human transitional cell carcinoma. Cancer Res 1993; 53: 5300-7.
- 180 Nicosia RF, Nicosia SV, Smith M. Vascular endothelial growth factor, platelet-derived growth factor, and insulinlike growth factor-1 promote rat aortic angiogenesis in vitro. Am J Pathol 1994; 145: 1023-9.
- 181 Goldman CK, Kim J, Wong WL, King V, Brock T, Gillespie GY. Epidermal growth factor stimulates vascular endothelial growth factor production by human malignant glioma cells: a model of glioblastoma multiforme pathophysiology. *Mol Biol Cell* 1993; 4: 121–33.
- 182 Fan TP, Hu DE, Guard S, Gresham GA, Watling KJ. Stimulation of angiogenesis by substance P and interleukin-1 in the rat and its inhibition by NK-1 or interleukin-1 receptor antagonists. *Br J Pharmacol* 1993; 110: 43-9. Wojta J, Gallicchio M, Zoellner H, Filonzi EL, Hamilton
- JA, McGrath K. Interleukin-4 stimulates expression of urokinase-type-plasminogen activator in cultured human foreskin microvascular endothelial cells. Blood 1993; 81: 3285-92
- 184 Smith DR, Polverini PJ, Kunkel SL, Orringer MB, Whyte RI, Burdick MD et al. Inhibition of interleukin 8 attenuates angiogenesis in bronchogenic carcinoma. J Exp Med 1994; 179: 1409-15.
- 185 Angiolillo AL, Kanegane H, Sgadari C, Reaman GH, Tosato G. Interleukin-15 promotes angiogenesis in vivo. Biochem Biophys Res Commun 1997; 233: 231-7.

- 186 Moenner M, Gusse M, Hatzi E, Badet J. The widespread expression of angiogenin in different human cells suggests a biological function not only related to angiogenesis. Eur J Biochem 1994; 226: 483-90.
- 187 Tsoplanglou PE, Pipili-Synetos E, Maragoudakis ME. Thrombin promotes angiogenesis by a mechanism independent of fibrin formation. Am J Physiol 1993; 264: C1302-7.
- 188 Cornelius LA, Nehring LC, Roby JD, Parks WC, Welgus HG. Human dermal microvascular endothelial cells produce matrix metalloproteinases in response to angiogenic factors and migration. J Invest Dermatol 1995; 105: 170-6.
- 189 Rosen EM, Zitnik RJ, Elias JA, Bhargava MM, Wines J, Goldberg ID. The interaction of HGF-SF with other cytokines in tumor invasion and angiogenesis. EXS 1993; 65: 301-10.
- 190 Bikfalvi A, Han ZC. Angiogenic factors are hematopoietic growth factors and vice versa. Leukemia 1994; 8: 523-9.
- Bussolino F, Arese M, Montrucchio G, Barra L, Primo L, Benelli R et al. Platelet activating factor produced in vitro by Kaposi's sarcoma cells induces and sustains in vivo angiogenesis. J Clin Invest 1995; 96: 940-52.
- 192 Jackson D, Volpert OV, Bouck N, Linzer DI. Stimulation and inhibition of angiogenesis by placental proliferin and proliferin-related protein. Science 1994; 266: 1581-4.
- 193 Magee JC, Stone AE, Oldham KT, Guice KS. Isolation, culture, and characterization of rat lung microvascular endothelial cells. Am J Physiol 1994; 267: LA33-641.
- 194 Ben-Av P, Crofford LJ, Wilder RL, Hla T. Induction of vascular endothelial growth factor expression in synovial fibroblasts by prostaglandin E and interleukin-1: a potential mechanism for inflammatory angiogenesis. FEBS Lett 1995; **372**: 83–7.
- 195 Hildenbrand R, Dilger I, Horlin A, Stutte HJ. Urokinase plasminogen activator induces angiogenesis and tumor vessel invasion in breast cancer. Pathol Res Pract 1995: 191: 403-9.
- 196 Canfield AE, Schor AM. Evidence that tenascin and thrombospondin-1 modulate sprouting of endothelial cells. J Cell Sci 1995; 108; 797–809.
- Ziche M, Morbidelli L, Masini E, Amerini S, Granger HJ, Maggi CA et al. Nitric oxide mediates angiogenesis in vivo and endothelial cell growth and migration in vitro promoted by substance P. J Clin Invest 1994; 94: 2036-44.
- 198 Mitchell CA, Davies MJ, Grounds MD, McGeachie JK, Crawford GJ, Hong Y et al. Enhancement of neovascularization in regenerating skeletal muscle by the sustained release of erucamide from a polymer matrix. J Biomater Appl 1996; 10: 230-49.
- 199 Kobayashi S, Okabe M, Kimura I, Kimura M. Interferongamma-activated macrophages release interleukin-1 alpha to increase tube formation from endothelial cells of rat aorta. Immunopharmacology 1995; 31: 93-101.
- 200 Angiolillo Al, Sgadari C, Tosato G. A role interferon-inducible protein 10 in inhibition of angiogenesis
- by interleukin-12. Ann NY Acad Sci 1996; 795: 158-67. 201 Vukanovic J, Isaacs JT. Linomide inhibits angiogenesis, growth, metastasis, and macrophage infiltration within rat

- prostatic cancers. Cancer Res 1995; 55: 1499-504.
- 202 Volpert OV, Ward WF, Lingen MW, Chesler L, Solt DB, Johnson MD et al. Captopril inhibits angiogenesis and slows the growth of experimental tumors in rats. J Clin Invest 1996; 98: 671-9.
- 203 Strieter RM, Polverini PJ, Arenberg DA, Walz A, Opdenakker G, Van Damme J et al. Role of C-X chemokines as regulators of langiogenesis in lung cancer. J Leukoc Biol 1995; 57; 752-62.
- 204 Millauer B, Longhi MP, Plate KH, Shawver LK, Risau W, Ullrich A et al. Dominant negative inhibition of Flk-1 suppresses the growth of many tumor types in vivo. Cancer Res 1996; 56: 1615-20.
- 205 Fotsis T, Zhang Y, Pepper MS, Aldercreutz H, Montesano R, Nawroth PP et al. The endogenous oestrogen metabolite 2-methoxyoestradiole inhibits angiogenesis and suppresses tumour growth. Nature 1994; 368: 237-9.
- 206 Takigawa M, Nishida Y, Suzuki F, Kishi J, Yamashita K, Hayakawa T. Induction of angiogenesis in chick yolk-sac membrane by polyamines and its inhibition by tissue inhibitors of metalloproteinases (TIMP and TIMP-2). Biochem Biophys Res Commun 1990; 171: 1264-71.
- 207 Pienta KJ, Nguyen NM, Lehr JE. Treatment of prostate cancer in the rat with the synthetic retinoid fenretinide. Cancer Res 1993; 53: 224-6.
- 208 Thurman GB, Page DL, Wamil BD, Wilkinson LE, Kasami M, Hellerqvist CG. Acute inflammatory changes in subcutaneous microtumors in the ears of mice induced by intravenous CM 101 (GBS toxin). J Cancer Res Clin Oncol 1996; 122; 549-53.
- 209 Nguyen NM, Lehr JE, Pienta KJ. Pentosan inhibits angiogenesis in vitro and suppresses prostate tumor growth in vivo. Anticancer Res 1993; 13: 2143-7.
- 210 Hu DE, Fan TP. Suppression of VEGF-induced angiogenesis by the protein tyrosine kinase inhibitor, lavendustin A. Br J Pharmacol 1995; 114: 262-8.
- 211 Hori Y, Hu DE, Yasui K, Smither RL, Gresham GA, Fan TP. Differential effects of angiostatic steroids and dexamethasone on angiogenesis and cytokine levels in rat sponge implants. *Br J Pharmacol* 1996; **118**: 1584–91. 212 Yamamoto T, Terada N, Nishizawa Y, Petrow V.
- Angiostatic activities of medroxyprogesterone acetate and its analogues. Int J Cancer 1994; 56: 393-9.
- 213 Yoon TJ, Yoo YC, Choi OB, Do MS, Khang TB, Lee SW et al. Inhibitory effect of Korean mistletoe (Viscum album coloratum) extract on tumour angiogenesis and metastasis of haematogenous and non-haematogenous tumour cells in mice. Cancer Lett 1995; 97: 83-91.
- 214 Tarnawski A, Hollander D, \$tachura J, Sarfeh IJ, Gergely H, Krause WJ et al. Angiogenic response of gastric mucosa ethanol injury is abolished by indomethacin. Gastroenterology 1989; 96: A505 (Abstract).
- 215 Lala PK, Al-Mutter N, Orucevic A. Effects of chronic indomethacin therapy on the development and progression of spontaneous mammary tumors in C3H/HEJ mice. Int J Cancer 1997; 73: 371-80.